

# Simulation Methods

```
#devtools::load_all()
suppressPackageStartupMessages({
  library(GEM)
  library(ggplot2)
  library(ggthemes)
  library(dplyr)
  library(tidyr)
  library(cowplot)
  library(magrittr)
})
```

## Run simulation

### Simulation Model

The simulation study was designed to evaluate the performance of our proposed method for analyzing aggregated binary outcomes. We generated data according to the following procedure:

### Data Generation Process

We simulated a dataset with  $n = 10,000$  candidate mutations with  $p = 10$  features/covariates, assigned to  $k = 100$  distinct samples/individuals. The data generation process follows these steps:

1. **Grouping Structure:** Mutations were randomly assigned to  $k = 100$  individuals, with each mutation having equal probability of assignment to any sample. Let  $\mathcal{S}_j$  represent the set of indices of mutations belonging to the  $j$ -th sample, for  $j \in \{1, 2, \dots, k\}$ .

2. **Covariate Generation:** We generated a covariate matrix  $\mathbf{X} \in \mathbb{R}^{n \times p}$  with each element independently drawn from a standard normal distribution:

$$X_{ij} \sim \mathcal{N}(0, 1) \quad \text{for } i = 1, \dots, n \text{ and } j = 1, \dots, p$$

3. **Mutation Status:** For each observation, a latent linear predictor  $\eta_i$  was computed as:

$$\eta_i = \mathbf{X}_i^T \beta + \beta_0$$

where:

- $\mathbf{X}_i$  is the vector of covariates for the  $i$ -th observation
- $\beta \in \mathbb{R}^p$  is a vector of regression coefficients, with each element independently drawn from  $\mathcal{N}(0, 2)$
- $\beta_0 = -0.8$  is a fixed intercept term

Binary mutation statuses  $Z_i$  were then generated according to a Bernoulli distribution with probability determined by the logistic function:

$$Z_i \sim \text{Bernoulli}(\text{logit}^{-1}(\eta_i)) \quad \text{for } i = 1, \dots, n$$

where  $\text{logit}^{-1}(x) = \frac{1}{1+e^{-x}}$  is the inverse logit (or logistic) function.

4. **Simulated age at blood draw:** For each sample  $j$ , we computed the true count of positive binary outcomes:

$$C_j = \sum_{i \in \mathcal{S}_j} Z_i \quad \text{for } j = 1, \dots, k$$

The final response variable (simulated age at blood draw)  $Y_j$  for each aggregation unit was generated as:

$$Y_j = 60 + 10 \cdot \log_2(C_j) \cdot \beta_1 + \epsilon_j \quad \text{for } j = 1, \dots, k$$

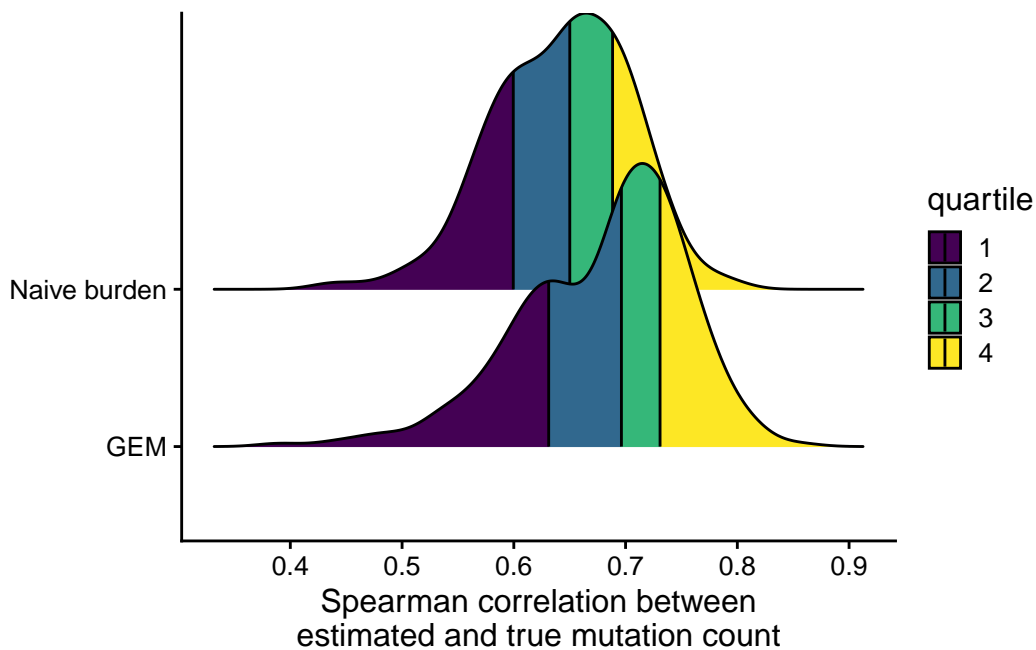
where:

- $\beta_1 = 1.0$  is a fixed coefficient
- $\epsilon_j \sim \mathcal{N}(0, 5)$  represents random noise
- The logarithmic transformation  $\log_2(C_j)$  models a non-linear relationship between counts and the response

We then run this simulation 300 times and estimate the concordance with the “true” mutation burden using both GEM and the naive mutation count/burden as estimators.

```
sims = GEM:::wrap_simulation(nsims = 300, .progress = FALSE)
```

```
sims %>%
  rename(
    `GEM` = cor_gem,
    `Naive burden` = cor_burden
  ) %>%
  tidyr::pivot_longer(cols = everything(), names_to = "type", values_to = "cor") %>%
  ggplot(., aes(y = type, x = cor, fill = factor(stat(quantile)))) +
  stat_density_ridges(
    geom = "density_ridges_gradient",
    calc_ecdf = TRUE,
    quantiles = 4,
    quantile_lines = TRUE
  ) +
  scale_fill_viridis_d(name = "quartile") +
  theme_cowplot(font_size = 12) +
  theme(axis.title.y = element_blank()) +
  labs(x = "Spearman correlation between\nestimated and true mutation count")
```



```
summary(sims)
#>      cor_gem      cor_burden
#> Min.   :0.3898  Min.   :0.4326
```

```
#> 1st Qu.:0.6312 1st Qu.:0.5995
#> Median :0.6963 Median :0.6501
#> Mean   :0.6797 Mean   :0.6434
#> 3rd Qu.:0.7309 3rd Qu.:0.6884
#> Max.    :0.8545 Max.    :0.7971
```